Stable Nitrogen and Carbon Isotope Ratios Indicate Traditional and Market Food Intake in an Indigenous Circumpolar Population

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Abstract

The transition of a society from traditional to market-based diets (termed the nutrition transition) has been associated with profound changes in culture and health. We are developing biomarkers to track the nutrition transition in the Yup’ik Eskimo population of Southwest Alaska based on naturally occurring variations in the relative abundances of carbon and nitrogen stable isotopes ($\delta^{15}N$ and $\delta^{13}C$ values). Here, we provide three pieces of evidence toward the validation of these biomarkers. First, we analyzed the $\delta^{15}N$ and $\delta^{13}C$ values of a comprehensive sample of Yup’ik foods. We found that $\delta^{15}N$ values were elevated in fish and marine mammals and that $\delta^{13}C$ values were elevated in market foods containing corn or sugar cane. Second, we evaluated the associations between RBC $\delta^{15}N$ and $\delta^{13}C$ values and self-reported measures of traditional and market food intake ($n = 230$). RBC $\delta^{15}N$ values were correlated with intake of fish and marine mammals ($r = 0.52; P < 0.0001$). RBC $\delta^{13}C$ values were correlated with intake of market foods made from corn and sugar cane ($r = 0.46; P < 0.0001$) and total market food intake ($r = 0.46; P < 0.0001$). Finally, we assessed whether stable isotope ratios captured population-level patterns of traditional and market intake ($n = 1003$). Isotopic biomarkers of traditional and market intake were associated with age, community location, sex, and cultural identity. Self-report methods showed variations by age and cultural identity only. Thus, stable isotopes show potential as biomarkers for monitoring dietary change in indigenous circumpolar populations.

Introduction

The transition of a society from traditional to market-based diets (termed the nutrition transition) has been associated with profound changes in culture and health (1–4). Many indigenous circumpolar populations are undergoing this transition (5–7), which is associated with increased rates of chronic disease (6, 8). Dietary change can be difficult to monitor due in part to the lack of baseline data and in part to the limitations of existing methods for dietary assessment. Self-report methods that are feasible to collect from large populations (e.g., FFQ) are subject to large error and bias (9, 10), whereas more reliable methods (e.g., repeated 24-h recall) can be prohibitively expensive (11, 12). Dietary biomarkers provide a promising alternative to self-report methods, because they are unbiased, more reliable, and can be measured from archived samples (13–16). We are developing biomarkers of traditional and market intake for the Yup’ik Eskimo population of Southwest Alaska based on comparisons of the marine fatty acids EPA and DHA (17, 18). Thus, we propose that $\delta^{15}N$ will indicate consumption of traditional marine foods in this population. The carbon isotope ratio ($\delta^{13}C$)
is elevated in plants using the C4 (Hatch-Slack) photosynthetic pathway relative to those using the more common C3 (Calvin) photosynthetic pathway (33). The most common representatives of these plants in the U.S. agricultural system are corn and sugar cane, which are widely present in the market diet as sweeteners (29), as ingredients in processed foods, and indirectly via domestic animals raised on corn (34,35). The carbon isotope ratio has shown moderate associations with reported C4-based sweetener and sweetened beverage intake in the U.S. population (23,24,36). Here, we propose that δ13C will provide an independent biomarker of market food intake for the Yup'ik Eskimo population.

The overall objective of this study was to evaluate isotopic biomarkers of market and traditional food intake in a Yup’ik Eskimo study population. Developing reliable and accurate markers of dietary change for this population could help to predict increases in disease incidence and develop appropriate dietary interventions. The specific aims of this study were 3-fold. First, we determined expected relationships between dietary intake and RBC isotope ratios by completing a comprehensive analysis of δ15N and δ13C values in traditional and market foods important to the Yup’ik population. Second, we evaluated the association between RBC δ15N and reported fish and marine mammal intake, and RBCδ13C and reported market food intake based on 4 d of diet records from 230 Yup’ik Eskimos. Finally, we evaluated whether variations in dietary intake by age, community location, and cultural identity that were previously reported for this population based on self-report were also seen using isotopic biomarkers (30,37–39). The extensive nature of previous dietary assessment in this population provides an ideal framework with which to evaluate the efficacy of these proposed biomarkers.

**Methodology**

**Participant recruitment and procedures.** Data are from the Center for Alaska Native Health Research study, a cross-sectional, community-based, participatory research study of genetic, nutritional, and psychosocial risk factors affecting obesity and related disease in Yup'ik Eskimos (40,41). The Center for Alaska Native Health Research study was approved by the University of Alaska Fairbanks Institutional Review Board, the National and Area Indian Health Service Institutional Review Boards, and the Yukon-Kuskokwim Health Corporation Human Studies Committee.

Between 2003 and 2005, 1003 participants aged 14–94 y were recruited from 10 communities in southwest Alaska as described in detail elsewhere (41). We categorized these communities as either coastal (<5 miles from the coast) or inland. At entry into the study, participants completed extensive interviewer-administered demographic questionnaires. A subset of 315 participants from the first 7 communities completed a single interviewer-administered 24-h dietary recall as well as a 3-d food record. A subset of 767 participants from all 10 communities completed a wellness questionnaire. We used the responses to two questions about cultural identification, which asked how much an individual felt they followed a Yup'ik or Kass'aq (non-native) way of life. Responses to these questions were coded numerically (1 = follows the lifestyle a lot, 2 = some, 3 = not at all) and were not mutually exclusive (e.g., a person could respond “follows the lifestyle a lot” for both ways of life).

**Biological sample collection.** Blood was collected into EDTA tubes and processed in rural communities using a portable centrifuge. Serum, lymphocytes, and RBC were stored in aliquots at −20°C in a portable freezer. Within 6 d, samples were shipped to the University of Alaska Fairbanks and stored at −80°C. Aliquots of RBC were removed for stable isotope analysis, as described below. RBC were chosen for analyses, because they reflect dietary intake over a period of 1–3 mo (42–44) and thus provide a more stable estimate of dietary intake than serum (45). RBC aliquots were autoclaved for 20 min at 121°C to destroy blood-borne pathogens, apportioned into tin capsules (3.5 × 3.75 mm), and oven dried at 60°C to a final weight of 0.2–0.4 mg. Neither autoclaving nor the use of EDTA-treated tubes affects RBC carbon or nitrogen isotope ratios (28).

**Food sample collection.** We sampled a broad range of traditional and market foods that were known to contribute to this study population’s diet (n = 254). Foods were defined based on their NDS-R (software version 4.06; University of Minnesota) food codes. Traditional foods were harvested from the local environment and samples were donated by residents from 3 Yup’ik Eskimo communities. Market foods were purchased from community grocery stores or grocery stores in Fairbanks, Alaska. We sampled 3 or more representatives of foods contributing >5% of energy (based on dietary self-report data), and one or more representatives from foods contributing 1–5% of energy. We sampled more rarely consumed traditional food items (contributing <1% energy) when donated. Marine mammal samples were collected under permit number 932-1905-00IMA-009526 issued by the National Marine Fisheries Service and the U.S. Fish and Wildlife Service, under the authority of the Marine Mammal Protection Act and Endangered Species Act. A list of foods sampled and their sampling frequencies is given in Supplemental Table 1.

Food samples were grouped into traditional or market-based food groups. Traditional foods were divided into four groups: aquatic (marine mammals, fish, and seal oil), terrestrial animals (birds and mammals), terrestrial plants (berries and wild plants), and waterfowl (ducks, geese, and swans). We defined waterfowl separately, because these species forage in both marine and terrestrial habitats. Furthermore, we note that the “aquatic” category contains both marine and freshwater fish species. Although freshwater and marine fish are expected to differ slightly in δ15N and δ13C values, their values are likely more similar to each other than to other classes of foods. Market foods were divided into five groups: market grains and vegetables (foods from C3 plants, including pasta, wheat, rice, nuts, fruits, and vegetables), corn and cane sugar (C4-based foods, including beverages sweetened with high fructose corn syrup, cane sugar, candy, corn), meat (chicken, turkey, beef, eggs), dairy (milk, cheese), or mixed (containing ingredients from more than one group). For isotope analysis, food samples were oven dried at 60°C for at least 48 h, ground to a fine powder, and weighed into tin capsules to a final weight of 0.2–0.4 mg.

**Stable isotope analysis.** Samples were analyzed at the Alaska Stable Isotope Facility by continuous-flow isotope ratio MS using a Costech ECS4010 Elemental Analyzer (Costech Scientific) interfaced to a Finnigan Delta Plus XP isotope ratio mass spectrometer via the Conflo III interface (Thermo-Finnigan). The conventional means of expressing natural abundance isotope ratios are as δ values in permil (‰) relative to international standards, defined as 6X = 1000 × ([Rsample/Rstandard]) − 1 (46). Here, R is the ratio of heavy to light isotope (15N/14N or 13C/12C) and the standards are atmospheric N for nitrogen and PeeDee Belemnite for carbon. Because the foods and RBC samples from this study had a lower 13C/12C than the standard, the values of δ13C were negative. We concurrently prepared and ran multiple peptone working standards (δ15N = 7.0, δ13C = −15.8‰, n = 128) to assess analytical accuracy and precision, measured as the SD of these analyses. Accuracy was within 0.1% and precision was within 0.2% for both isotopes.

**Assessment of dietary intake.** Dietary intake was estimated using an interviewer-administered 24-h dietary recall and a 3-d food record. Data from these instruments were combined to achieve a stable estimate of dietary intake. The 24-h dietary recall was collected from each participant by certified interviewers using a computer-assisted recall (NDS-R version 4.06). Participants were asked to recall all food and beverages consumed over the 24-h period covering the day prior to the interview using a multiple pass approach to minimize recall bias. Although most participants were bilingual, a native Yup’ik speaker trained in the use of NDS-R was available for non-English speakers.

When completing the 3-d food record, participants were instructed to maintain their usual eating habits. A research team member reviewed all...
records for completeness, which were then entered into the NDS-R software package by certified coders. A second reviewer researched all entries for accuracy. Records were considered unreliable and excluded from analysis if daily energy intake was >5000 or <500 kcal (38 participants had 1 d excluded, 4 had 2 d excluded). Individuals who had >2 d considered unreliable (\( n = 2 \)) or whose 3-d food record or 24-h recall was incomplete (\( n = 83 \)) were excluded from self-report analyses; 230 individuals remained.

**Dietary analysis.** The contributions of traditional and market foods to an individual’s diet were assessed as follows: all food items were assigned to traditional and market food groups (as defined above and given in Supplemental Table 1) based on their food codes from the NDS-R Food and Nutrient Database 33 (July 2003). A few Alaska Native foods were missing from the database; these were substituted for similar food items where appropriate or the foods were added to the database. We then summed total energy consumed for 3 categories of foods: traditional aquatic, market, and \( C_4 \)-based market foods and used these totals for summing total energy consumed for 3 categories of foods: traditional aquatic, market, and \( C_4 \)-based market foods and used these totals for dietary analysis.

Because market foods included food groups that were partially \( C_4 \) based (mixed foods, meats, and dairy) as well as entirely \( C_4 \) based (corn and cane sugar), we defined \( C_4 \)-based market food intake by weighting isotope ratios, and a multiple linear regression model including demographic variables (49). Agreement between the two methods was good (\( R^2 = 0.78 \)). This fractional content (\( f_{C4} \)) was calculated using the mean \( \delta^{13}C \) of food groups and an isotopic mixing model, as follows:

\[
f_{C4} = \frac{(\delta^{13}C_{food \ group \ mean} - \delta^{13}C_{C4})}{(\delta^{13}C_{C4} - \delta^{13}C_{C3})},
\]

where \( f_{C4} \) is the fraction of the food that is \( C_4 \) based, \( \delta^{13}C_{C4} \) is the mean carbon isotope ratio of \( C_4 \)-based plant foods (corn and cane sugar) (Table 1), and \( \delta^{13}C_{C3} \) is the carbon isotope ratio of \( C_3 \)-based plant foods (grains and vegetables (47)). We note that this is a highly simplified mixing model that does not take into account differences in macronutrient composition among foods (48) and uses a mean \( \delta^{13}C \) for food classes rather than adjusting foods individually. However, the purpose of the calculation is to get a broad estimate of how much of the market diet is derived from \( C_4 \) sources rather than to present a highly precise measure.

**Correcting RBC \( \delta^{13}C \) values for the influence of fish and marine mammal intake.** Our aim was to use \( \delta^{13}C \) as an index of \( C_4 \)-based market food intake; however, both \( C_4 \) and aquatic foods have elevated \( \delta^{13}C \) values relative to \( C_3 \)-based market foods (27,29). Therefore, we used \( \delta^{13}N \) values to adjust for the influence of aquatic foods on RBC \( \delta^{13}C \) as follows:

\[
\delta^{13}C_{total} = \delta^{13}C_{fish} + \frac{\delta^{13}C_{water}}{15N_{water}} \times \frac{\delta^{13}N_{water}}{\delta^{15}N_{water}},
\]

where \( \delta^{13}C_{total} \) is adjusted \( \delta^{13}C \) value, \( \delta^{13}C \) and \( \delta^{13}N \) are measured RBC isotope ratios, and \( \delta^{13}N_{water} \) is mean \( \delta^{13}N \) value for all members of the population reporting no traditional aquatic food intake (\( n = 42 \)). This was measured to be 7.8‰, \( \delta^{13}N_{water} / \delta^{15}N_{water} = \delta^{13}C_{total} / \delta^{13}C_{fish} \) for each unit increase in \( \delta^{13}C \) across all fish and marine mammal samples. This was measured to be 0.91 (see Results).

We tested the accuracy of this approach by assessing the agreement between prediction of \( C_4 \)-based market foods based on \( \delta^{13}C \) values and a multiple linear regression model including \( \delta^{13}C \) and \( \delta^{15}N \) as independent variables (49). Agreement between the two methods was good (mean difference: 0.0 ± 1.7% of total energy). Although both of these methods account for the influence of aquatic food intake on \( \delta^{13}C \) values, we chose to adjust RBC \( \delta^{13}C \) values in this study to generate a single, independent variable that can be used in multiple analyses.

**Statistical analyses.** All statistical analyses were performed using JMP version 8 (SAS Institute). We evaluated differences in the sex and age distributions between the complete sample of participants and those who completed dietary interviews or cultural identity questions using a chi-square test. We used ANOVA to compare the isotope ratios of food samples in each market and traditional food group and compared means using Tukey-Kramer’s honest significant difference. We assessed the associations between isotope ratios and dietary intake data, as well as between dietary intake variables, using linear regression.

**Results**

**Stable isotope ratios of food.** Nitrogen isotope ratios varied among traditional and market food groups (\( \delta^{15}N; P < 0.001 \)) (Table 1; Supplemental Table 1). Traditional aquatic foods had substantially higher \( \delta^{15}N \) values than any other food group (Table 1). Marine fish species had higher \( \delta^{15}N \) values than freshwater fish species (\( P = 0.0013 \), marine \( \delta^{15}N = 14.3 ± 3.4‰ \), freshwater \( \delta^{15}N = 11.5 ± 2.7‰ \)); however, their values overlapped and were both significantly higher than all other food groups. Thus, we continued to group these species together as “aquatic foods” for further analyses. Excluding aquatic foods, animal-based food groups had higher mean \( \delta^{15}N \) values than plant-based food groups (\( P = 0.0037 \)). We found a strong positive relationship between \( \delta^{13}C \) and \( \delta^{15}N \) values of tradi-

<table>
<thead>
<tr>
<th>Table 1</th>
<th>Nitrogen and carbon isotope ratios of market and traditional foods</th>
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</thead>
<tbody>
<tr>
<td><strong>Food group</strong></td>
<td><strong>( \delta^{15}N(%) )</strong></td>
</tr>
<tr>
<td><strong>Subsistence foods</strong></td>
<td></td>
</tr>
<tr>
<td>Aquatic</td>
<td>14.2 ± 3.2a</td>
</tr>
<tr>
<td>Waterfowl</td>
<td>7.3 ± 2.1b</td>
</tr>
<tr>
<td>Terrestrial animals</td>
<td>3.7 ± 4.2c</td>
</tr>
<tr>
<td>Terrestrial plants</td>
<td>-0.3 ± 2.2</td>
</tr>
<tr>
<td><strong>Market foods</strong></td>
<td></td>
</tr>
<tr>
<td>Corn and cane sugar</td>
<td>4.0 ± 0.8c</td>
</tr>
<tr>
<td>Meat</td>
<td>3.7 ± 1.4d</td>
</tr>
<tr>
<td>Mixed</td>
<td>1.9 ± 2.3c</td>
</tr>
<tr>
<td>Dairy</td>
<td>4.5 ± 0.5d</td>
</tr>
<tr>
<td>Grains and vegetables</td>
<td>2.1 ± 1.9c</td>
</tr>
</tbody>
</table>

1 Values are mean ± SD. Means in a column without a common letter differ, \( P < 0.05 \). Means were calculated using the mean isotope ratio for each food within the group, given in Supplemental Table 1.
2 Carbon isotope ratios are negative, because all samples had less \( 13C \) than the standard against which they were measured.
3 Includes marine mammals and marine and freshwater fish species.
tional foods ($\beta = 1.86; r = 0.82; P < 0.0001$) (Fig. 1A) and aquatic foods only ($\beta = 0.91; r = 0.71; P < 0.0001$) but not market foods ($\beta = 0.14; r = 0.38; P = 0.08$) (Fig. 1B).

Carbon stable isotope ratios differed between food groups ($\delta^{13}C$: $P < 0.001$) (Fig. 1), with a clear distinction between $C_3$ and $C_4$-based foods. Corn and cane sugar ($C_4$) had elevated $\delta^{13}C$ values relative to grains and vegetables ($C_3$) (Table 1). The mean isotopic difference between these groups was almost 13‰ and was consistent with reported $\delta^{13}C$ values for $C_3$ and $C_4$ plants generally ($P < 0.0001$; marine $\delta^{13}C = -20.4 \pm 1.8$‰, freshwater $\delta^{13}C = -24.3 \pm 3.3$).

Sample population. Females were slightly over-represented relative to males in the whole study population (54%), the subset of participants with dietary self-report data (59%), and the subset of participants reporting cultural identification (56%; all $P < 0.01$) (Table 2). The age distribution of participants reporting cultural identification did not differ from the complete study sample ($P < 0.01$), with reduced participation by those who were >60 y old. The age distribution of participants reporting cultural identification did not differ from the complete study sample.

Associations between stable isotope ratios and reported dietary intakes. Nitrogen isotope ratios were significantly correlated with intake of traditional aquatic foods based on dietary self-report (Table 3). Reported intake of total traditional and traditional aquatic foods was highly correlated (Table 3). Aquatic foods accounted for 77% energy from traditional sources.

Adjusted carbon isotope ratios were positively correlated with $C_4$ market food intake (Table 3) as well as total market food intake. Intake of $C_4$-based market foods was positively associated with total market food intake (Table 3). $C_4$ sources accounted for 40% of total energy from market foods.

Population level patterns of traditional aquatic food intake. Mean RBC $\delta^{15}N$ values reflected a high and variable intake of aquatic foods (Table 2). Aquatic foods were reported by 82% of people with dietary self-report data ($n = 188$) and mean intake was $210 \pm 245$ kcal.

We found strong associations between RBC $\delta^{15}N$ values and sex, community location, and age (Table 2). Values of RBC $\delta^{15}N$ increased with age ($\beta = 0.052$; CI = 0.048–0.056; $P < 0.0001$) and were higher in coastal communities ($P < 0.0001$) and females ($P < 0.0001$). There was a community location by age interaction ($P < 0.0001$), because differences in $\delta^{15}N$ values between coastal and inland locations increased with age. In the self-report data, we found an increase in intake of aquatic foods with age ($\beta = 1.003; CI = 1.002–1.004; P < 0.0001$) but not community location ($P = 0.11$) or sex ($P = 0.09$).

Intake of aquatic foods was positively associated with following a Yup'ik way of life as indicated by nitrogen isotope ratios ($P < 0.0001$) and self-report data ($P = 0.016$). Following a Kass’aq way of life was negatively associated with nitrogen isotope ratios ($P = 0.045$) and self-reported intake of aquatic foods ($P = 0.045$).

Population level patterns of $C_4$-based market food intake. Adjusted RBC carbon isotope ratios reflected a mixed diet of $C_3$ and $C_4$ foods (Table 2). All participants for whom we have dietary self-report information reported consuming market foods ($n = 230$) and mean intake of $C_4$-based market foods was $543 \pm 301$ kcal.

We found strong associations between sex, age, and community location and RBC $\delta^{13}C_A$ values (Table 2). Values of RBC $\delta^{13}C_A$ decreased with age ($\beta = -0.057; CI = -0.062 to -0.053; P < 0.0001$), and were higher in upriver communities ($P < 0.0001$) and males ($P = 0.0011$). There was a community location by age interaction ($P = 0.0223$). In the self-report data, we did not find differences in market food intake by sex or community location, although there was a decrease in intake of market foods with age ($\beta = 0.98; P < 0.0001$) (Table 2).

RBC $\delta^{13}C_A$ values were positively associated with following a Kass’aq way of life ($P < 0.0001$) and negatively associated with following a Yup’ik way of life ($P < 0.0001$) (Table 2). Self-reported intake of $C_4$-based market foods was negatively associated with a Yup’ik way of life ($P = 0.0006$) (Table 2) but was not associated with a Kass’aq way of life ($P = 0.26$).

Discussion

Carbon and nitrogen stable isotope biomarkers capture patterns of traditional aquatic and market food intake in a Yup’ik Eskimo study population. Foods commonly consumed by the Yup’ik population exhibited highly distinct patterns of carbon and nitrogen isotope ratios, with elevated $\delta^{15}N$ and $\delta^{13}C$ values found in key traditional and market foods, respectively. Self-reported intake of these food groups was associated with RBC


### TABLE 2

<table>
<thead>
<tr>
<th>Isotope measures</th>
<th>Self-report measures</th>
</tr>
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<tbody>
<tr>
<td></td>
<td>$n$</td>
</tr>
<tr>
<td>Whole population</td>
<td>1003</td>
</tr>
<tr>
<td>Total</td>
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</tr>
<tr>
<td>Sex</td>
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<tr>
<td>Male</td>
<td>460</td>
</tr>
<tr>
<td>Female</td>
<td>543</td>
</tr>
<tr>
<td>Age, y</td>
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</tr>
<tr>
<td>14−&lt;20</td>
<td>200</td>
</tr>
<tr>
<td>21−&lt;40</td>
<td>374</td>
</tr>
<tr>
<td>41−&lt;60</td>
<td>303</td>
</tr>
<tr>
<td>&gt;60</td>
<td>126</td>
</tr>
<tr>
<td>Location, %</td>
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<tr>
<td>Coastal</td>
<td>402</td>
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<tr>
<td>Upriver</td>
<td>601</td>
</tr>
<tr>
<td>Cultural identifi-</td>
<td>767</td>
</tr>
<tr>
<td>cation subset</td>
<td></td>
</tr>
<tr>
<td>Yup’ik</td>
<td></td>
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<tr>
<td>High</td>
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<tr>
<td>Medium</td>
<td>399</td>
</tr>
<tr>
<td>Low</td>
<td>19</td>
</tr>
<tr>
<td>Kass’aq</td>
<td></td>
</tr>
<tr>
<td>High</td>
<td>129</td>
</tr>
<tr>
<td>Medium</td>
<td>575</td>
</tr>
<tr>
<td>Low</td>
<td>63</td>
</tr>
</tbody>
</table>

1 Values are mean ± SD.

2 Carbon isotope ratios are negative, because all samples had less $^{13}C$ than the standard against which they were measured.

$\delta^{15}N$ and $\delta^{13}C_A$ values. These isotope ratios detected demographic and cultural variations in both traditional and market food intake known to exist within this population (30) as well as some that were undetected by self-reported measures. Stable isotope markers have the potential to be useful in assessing the health impacts of dietary change in indigenous circumpolar populations, because they impose low participant burden and can be measured with precise, high throughput, and inexpensive methods.

Traditional aquatic foods had significantly elevated $\delta^{15}N$ values relative to all other food groups, causing a 1% increase in RBC $\delta^{15}N$ for each 5% increase in energy intake from aquatic foods. RBC $\delta^{15}N$ measurements showed that aquatic food consumption increased with age, reflecting trends in total traditional food intake described for this study population (30) and other circumpolar populations (38,52,53). RBC $\delta^{13}N$ values also captured differences in traditional aquatic food intake between coastal and inland communities, an effect seen primarily in elders, and a slight increase of aquatic food intake in women relative to men. Self-reported intake of traditional aquatic (this study) or total traditional foods (30) did not vary with community location or sex in this population. Thus, stable isotope biomarkers were able to identify patterns of traditional food intake that were not evident in dietary self-report data.

Corn and cane sugar-based market foods exhibited uniquely high $\delta^{13}C$ values relative to all other foods. Animal-based market foods (beef, pork, poultry, eggs, and dairy) also had elevated $\delta^{13}C$ values, reflecting corn in the diet of commercially raised animals (34,35). We refer collectively to these foods as “$C_4$-based market foods.” RBC $\delta^{13}C_A$ values were associated with intake of $C_4$-based market foods as well as total market food intake with each 8% increase in energy from market foods causing a 1% increase in $\delta^{13}C_A$. The carbon isotope ratio has been moderately associated with sweetener intake in other U.S.

### TABLE 3

<table>
<thead>
<tr>
<th>Relationships between stable isotope biomarkers and self-reported measures of traditional aquatic and $C_4$-based market food intake ($n=230$)</th>
</tr>
</thead>
<tbody>
<tr>
<td>$\delta^{15}N$ vs. total aquatic, %</td>
</tr>
<tr>
<td>Total aquatic vs. total traditional, kcal</td>
</tr>
<tr>
<td>$\delta^{13}C_A$ vs. $C_4$, %</td>
</tr>
<tr>
<td>$\delta^{13}C_A$ vs. total market, %</td>
</tr>
<tr>
<td>$C_4$ vs. total market, kcal</td>
</tr>
</tbody>
</table>

1 Independent variables are listed first.

2 Where the dependent variables have been transformed, estimates of $\beta$ are back-transformed for ease of interpretation.
populations (23,24,36); however, the development of this marker is complicated by its concurrent association with animal protein intake (24,54). In our study, the fact that multiple market foods are influenced by corn likely strengthens the association between δ13C and market food intake.

Although market foods are known to be a significant source of energy to all age groups in this study population (39), both self-report and isotope data showed that consumption of market foods decreased with age. Adjusted δ13C values also showed that coastal communities consumed slightly less market foods than those inland and men consumed slightly more market foods than women. Both RBC δ13CA and self-reported measures of market food intake were positively associated with a non-native (Kass'aaq) way of life. As expected, these patterns are the reverse of those found for traditional food intake. However, the use of δ13CA as an independent biomarker for market food intake based on sugar cane and corn provides an alternative way to assess the nutrition transition that, unlike δ15N, does not require traditional food intake to be aquatic. Such a biomarker could be particularly useful for assessing dietary change in Alaska Native populations relying more heavily on traditional foods such as mOOSE and caribou (55,56), which are not distinct from market foods in δ13N.

The primary limitation of this study was that we compared stable isotope ratios to self-reported estimates of traditional and market food intake, which are subject to error and biases associated with age, sex, and other individual characteristics. These errors may have obscured the true relationships between isotopic markers and diet in our study population. For example, relationships between δ13N and EPA were very strong for this population [r ≥ 0.8; (17,18)] compared with the relationship between δ13N and reported aquatic food intake presented here [r = 0.52]. However, this study had several unique strengths. The Yup'ik Eskimo population of Southwest Alaska is culturally and linguistically among the most intact of Alaska Native peoples and traditional food use is still common (41); therefore, this is an ideal population in which to test markers of both traditional and market foods. Furthermore, the extensive nature of dietary research in this population provided an ideal opportunity to identify relevant foods for isotopic sampling and test stable isotope biomarkers within the context of known dietary patterns.

In summary, we demonstrated that stable nitrogen and carbon isotope ratios in RBC indicate the use of traditional and market foods in a Yup'ik Eskimo study population in Southwest Alaska. Isotopic biomarkers have great potential due to their affordability, reliability, and ability to be measured noninvasively and with low burden (17,18). Furthermore, because these markers can be measured in stored specimens, they have the potential to provide baseline data for studies of dietary change over time (16). The development of reliable biomarkers of traditional and market-based food intake will help in the evaluation of the overall health impacts of dietary change in circumpolar populations.

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Literature Cited


