Cortical and Trabecular Bone, Bone Mineral Density, and Resistance to ex Vivo Fracture Are Not Altered in Response to Life-Long Vitamin A Supplementation in Aging Rats

Amanda E. Wray, Nori Okita, and A. Catharine Ross

Abstract

High vitamin A (VA) intakes have been correlated with increased risk of bone fracture. Over 50% of the U.S. adult population reports use of dietary supplements, which can result in VA intakes greater than 200% of the RDA. In this study, 2 experiments were designed to determine the effect of dietary VA on cortical and trabecular bone properties and resistance to ex vivo fracture. In Expt. 1, we investigated whether orally administered VA accumulates in bone. Seven-week-old rats were treated daily with VA (6 mg/d for 14 d). Total retinol increased in both the tibia and femur ($P < 0.01$). In Expt. 2, we conducted a longitudinal study in which rats were fed 1 of 3 levels of dietary VA (marginal, adequate, and supplemented, equal to 0.35, 4, and 50 $\mu$g retinol/g diet, respectively) from weaning until the ages of 2–3 month (young), 8–10 month (middle-aged), and 18–20 month (old). Tibial trabecular and cortical bone structure, bone mineral density, and resistance to fracture were measured using micro-computed tomography and material testing system analysis, respectively. The VA-marginal diet affected measures of cortical bone dimension, suggesting bone remodeling was altered. VA supplementation increased medullary area and decreased cortical thickness in young rats ($P < 0.05$), but these changes were not present during aging. VA supplementation did not affect resistance to fracture or bone mineral content in old rats. From these results, we conclude that VA-marginal status affects trabecular bone more than cortical bone, and VA supplementation at a moderate level over the lifetime is unlikely to increase the risk of age-related bone fracture in rats. J. Nutr. 141: 660–666, 2011.

Introduction

Vitamin A (VA) is a fat-soluble dietary component required for vision, cellular differentiation, proliferation, normal growth, and bone development (1,2). The principal biologically active metabolite of VA, retinoic acid, acts through retinoid-specific receptors to modulate gene expression (1). Both VA deficiency and toxicity (hypervitaminosis A) result in skeletal abnormalities. Spontaneous fractures are often noted as a well-known sign of hypervitaminosis A (3). However, hypervitaminosis A has most often been studied in the context of short-term animal studies using high doses of VA or case studies of humans who inadvertently ingested very high doses of VA, generally over a short period of time. The consequences of marginal intakes, or conversely, of moderately high but nontoxic intakes of VA over the lifetime are not understood.

For humans, the RDA for adult males and females is 900 and 700 $\mu$g retinol activity equivalents (RAE)/d, respectively, and the tolerable upper intake level is 3000 $\mu$g/d of preformed retinol (4). According to data from the 1999–2000 NHANES, 52% of U.S. adults report using a dietary supplement (5). Epidemiological studies report total dietary retinol intakes as high as 8771 $\mu$g/d RAE (6) and mean intakes of 1250 $\mu$g/d RAE are common (7–12).

As a fat-soluble vitamin, VA is absorbed and transported with TG and cholesterol in chylomicrons. After chylomicron TG undergo lipolysis in capillaries, chylomicron remnants then bind, primarily, to the hepatic apoprotein receptor, LDL receptor-related protein, LRP (2,12), and deliver most of their VA to the liver. However, bone marrow osteoblasts express LRP5 and have been reported to be the second largest clearance site for chylomicron remnants (13–16). Studies in rabbits using $^3$H-retinol as a tracer showed that a portion of newly absorbed chylomicron VA resided, at least transiently, in bone marrow in the postprandial period (13–16). Other studies have indicated uptake of chylomicron lipids by osteoblasts (15). Thus, the...
hypothesis that dietary VA accumulates in bone is plausible, and if VA weakens bone, it could lead over time to reduced bone strength.

Many nutrients affect the integrity of bone (17). Information in the literature is mixed regarding the response of bone to alterations in dietary VA. Studies of both self-reported high VA intake and experimental dosing have reported a variety of outcomes ranging from no interaction, a detrimental interaction, to a beneficial result (6–9,16,18–30). Elevated VA achieved through the use of supplements might be associated with negative consequences related to the risk of bone fracture and decreases in mineralization, although this too is controversial (10,20,25). Animal studies on VA and bone have typically been conducted over a short time period and have investigated only the period of rapid growth and/or have used pharmacologic doses of VA or synthetic retinoids known to be more toxic than VA (3,31,32). In the present study, we conducted 2 experiments to determine, first, whether VA accumulates in long bones after an acute exposure to high-dose VA and second, whether a range of intake of dietary VA, when consumed over the lifetime, affects the properties of cortical and trabecular bone and the bone’s ability to resist fracture. For the latter objectives, we used the tibia of rats that had been fed VA-marginal, -adequate, or -supplemented diets from weaning until age 2–3, 8–10, or 18–20 mo (33,34), thus allowing an assessment of the effects of diet on each measured variable at 3 times that are representative of the periods of bone growth (young rats), maintenance (middle-aged rats), and age-related bone loss (old rats), respectively.

Materials and Methods

Rats and diets. Approval for animal studies was obtained from the Institutional Animal Use and Care Committee of the Pennsylvania State University. Animals were housed under a normal 12-h-light/12-h-dark cycle in a climate controlled environment and had free access to food and water. In Expt. 1 (acute treatment), young male and female Sprague-Dawley rats obtained through mating within our facility were used to determine whole bone VA (measured as total retinol) content. In preparation, rat pups were fed a VA-deficient diet based on the AIN-93 Growth diet, prepared by Research Diets; this diet has been shown to reduce VA storage in nursing pups (35,36). After weaning at 25 d of age, the young rats continued to receive the same diet, while they were randomly assigned to receive either a daily oral supplement of all-trans retinyl palmitate [n = 5; 6 mg of retinol as retinyl palmitate (Sigma-Aldrich) dispersed in 100 μL canola oil daily] for 14 d or the same volume of oil alone as a control (n = 4). After treatment for 14 d, the rats were euthanized by carbon dioxide asphyxiation.

In Expt. 2 we evaluated the effects of VA status over the life span, from young to middle age to old age, on bone properties. This study utilized the tibia of male Lewis rats that had been obtained in a previously reported study (33,34). Briefly, rat pups were weaned as described above and randomly assigned in a 3 × 3 design to an AIN-93 purified diet, modified to contain 1 of 3 levels of VA (33,34,37): 0.35 (VA-marginal), 4.0 (VA-adequate level, designated as control), or 50 mg (VA-supplemented) retinol equivalents/kg diet. Diets were prepared by Dyets, Inc. and the VA was added as retinyl palmitate. The diets were continuously fed until the rats were 2–3, 8–10, or 18–20 mo old.

At the time of euthanasia and bone collection, the right tibia from each rat was removed and cleaned of excess tissue while submersed in ice-cold PBS. After thoroughly cleaning, they were stored individually at −20°C in sealed Whirl-pak containers until they were analyzed in the present Expt. 2. The number of rats for which tibiae (one tibia per rat) were available for analysis equaled, for young rats, n = 6, 9, 6; for middle-aged rats, 7, 6, 9; and for old rats, 8, 7, 6 for the VA-marginal, control, and VA-supplemented diets, respectively. Body weight was determined for the rats used in these analyses. For some analyses, e.g., to test breaking strength, a few bones that had broken were not suitable for analysis. The final n per assay is included in each figure.

Retinoid analysis. Whole bone total VA was determined as total retinol after extraction of the right tibia and femur from each VA-treated rat in Expt. 1. The wet weight was determined, followed by Folch extraction of total lipids (38). The washed Folch extract was evaporated under argon, saponified, and analyzed for total retinol as previously described (39). Plasma from the same rats was similarly analyzed. The extracts were dried and then reconstituted in methanol for analysis of total retinol content by reverse-phase HPLC at a wavelength of 325 nm (39).

Bone dimensional property measurements. In Expt. 2, bone dimensions, trabecular and cortical measurements, and mechanical testing were determined on tibia that had been stored as described above. Samples were rehydrated by complete submersion in 10 mL of sterile PBS at 4°C overnight. Preliminary studies using freshly collected tibia from VA-adequate rats were conducted to ensure that results were similar for the stored bones. For caliper measurements, the operator was unaware of each sample’s treatment group to prevent bias. Caliper (Traceable Digital Calipers, VWR) measurements were taken for total length, epiphyseal, sagittal, and coronal widths to detect changes in bone size or shape.

Micro-computed tomography (μCT) was used to reconstruct trabecular structure and measure periosteal and endosteal radii, cortical thickness, bone mineral density (BMD), number of trabeculae, trabecular thickness, and bone fraction. The proximal epiphysis was scanned using a desktop μCT system (μCT 40; Scanco Medical) at 15-μm voxel resolution with medium energy level (55 kVp and 145 μA) and 200-ms integration time.

A 5-mm section at the proximal metaphysis of the right tibia was scanned for histomorphological analyses by using the IPL scripts (Scanco Medical). An additional section was scanned for mid-diaphyseal shaft cross-sectional analysis using a purpose-written MATLAB program (version 6.5; MathWorks), as described previously (40). A group of 4 bone samples of similar length was scanned in a custom-built 30-mm cartridge made of poly methyl methacrylate to scan the same region of the bones.

Material testing system analysis. After scanning, tibiae were stored in PBS for no longer than 48 h at 4°C. Tibial 3-point flexural bending tests were conducted using a material testing apparatus (MiniBionix 858; MTS) with a 16-mm support span attached to a 22.7-g (50-lb) load cell (40), and crosshead speed was set at 2 mm/min. Bones were consistently oriented with the load applied on the posterior side at the midpoint of the midshaft, producing compression on the posterior surface and tension on the anterior surface. Load-displacement curve was obtained to determine force and displacement at yield and ultimate points of each tibia. These data along with the cross-sectional data at the midshaft were used to determine material properties. Stress (σ) and flexural modulus (E) were calculated by using $e = FL/cI$ and $E = F L^3/48ID$, respectively (40,41), where (F) was yield/ultimate loads in Newtons, (L) was the length of the support span in millimeters, (c) was the distance from the centroid to the periosteal tensile surface in millimeters, (l) was cross-sectional area moment of inertia about the mediolateral axis of the tibia in millimeters, and (d) was the displacement at the yield point in millimeters.

Ashing. After breaking, bones were collected for analysis. The samples were weighed to the 5th decimal place under standardized conditions, dried in a vacuum oven at 200°C for 24 h, cooled, reweighed to determine dry weight, and then ashed in a muffle furnace at 800°C for 24 h (40). Calculations were made according to the following formulas: water (%) = 100 (wet weight − dry weight)/wet weight; organic (%) = 100 (dry weight − ash weight)/wet weight; ash (%) = 100 (ash weight)/wet weight; tissue mineral (%) = 100 (ash weight/dry weight).

Statistical analysis. Data are presented as the mean ± SEM unless otherwise noted. Results in Expt. 1 were analyzed by 1-way ANOVA. Results in Expt. 2 were analyzed by 2-way ANOVA and P-values for the main effects of diet and age and their interaction are reported. The effect
of the level of dietary VA within each age group was further analyzed by least squares means test (SuperAnova). Values of $P < 0.05$ were considered significant.

**Results**

**Plasma and bone retinoid content after acute VA treatment in young rats.** In Expt. 1, plasma total retinol was significantly elevated in rats treated orally with VA for 14 d compared with oil-treated controls (Fig. 1A).

In the same rats, bone VA was significantly increased in both tibia and femur (Fig. 1B). The concentration was 25% higher in the tibia than in the femur of VA-treated rats. Short-term treatment with VA did not affect body or bone weights. Thus, Expt. 1 showed that ingestion of supplementary VA can increase the retinol content of bone and supported the use of the tibia for analysis of the effects of VA on bone properties.

**Dietary VA intake and body weight during aging.** In Expt. 2, we analyzed various bone variables in the tibia of male Lewis rats that had been fed VA-marginal, control, or -supplemented diets until the ages of 2–3, 8–10, or 18–20-mo, respectively. Age was a main factor ($P < 0.0001$; Table 1), as expected for a model of growth and aging (33,34), whereas diet had no effect on body weight until old age, when VA-marginal and -supplemented rats differed from each other, but neither differed from the control group.

**Mid-diaphyseal bone medullary area, cortical thickness, and endosteal and periosteal radii.** The mid-diaphyseal medullary area of the tibia, cortical thickness, endosteal radii, and periosteal radii were determined as indicators of bone growth and shape change (Table 1). Age was a factor for each variable ($P < 0.0001$). The medullary area of the tibia was smaller in VA-supplemented compared with VA-marginal rats (middle age and old) and control rats (old group only; $P < 0.05$)

![FIGURE 1](https://example.com/figure1.png)  
**FIGURE 1** Plasma retinol ([A]) and long-bone total retinol ([B]) content in tibia and femur of VA- and placebo (oil) treated rats. Values are means ± SEM, n = 4 (control) or 5 (VA treated) per bone. Means without a common letter differ, $P < 0.05$.

To determine whether the change in medullary area was due to alterations in the endosteal (inner) or periosteal (outer) bone surfaces, the radius from the centroid to each of the locations above was determined using MATLAB software (Table 1; schematic of measurements in Supplemental Fig. 2). There was an age-related increase in endosteal radius in old compared with young and middle-age rats ($P < 0.05$). Endosteal radius was larger in VA-marginal than supplemented rats (middle-age and old) ($P < 0.05$) (Table 1). To ensure that these differences were not compounded with a change in bone shape, the periosteal radius was also measured (Table 1). By middle and old age, periosteal radius was greater in VA-marginal compared with supplemented rats ($P < 0.05$), similar to medullary area and endosteal radius. Thus, bone dimensions tended to change in proportion to one another during aging, with VA-marginal > control > supplemented for old rats for medullary area and periosteal radius, and VA-marginal > control supplemented rats for endosteal radius.

Because there was a difference in endosteal and/or periosteal radii with diet during aging, we also determined mid-diaphyseal cortical thickness. Mid-diaphyseal cortical bone thickness differed with age ($P < 0.0001$), being greatest in middle-age rats. Differences due to diet were observed between the young control group and both the VA-marginal and VA-supplemented groups ($P < 0.05$) but not in other age groups.

**Trabecular number, thickness, and BMD.** Trabecular number decreased with age, as expected (Table 2). The tibia of old VA-supplemented rats contained more trabeculae per area compared with old VA-marginal rats ($P < 0.05$). Conversely, trabecular thickness increased with age (Table 2). Trabeculae were thicker in middle-aged and old VA-marginal rats compared with control and VA-supplemented rats of the same ages ($P < 0.05$). Trabecular BMD, determined by $\mu$CT, also increased with age and showed small but significant differences with intake of VA, similar to trabecular thickness (Table 2).

**Resistance of bone to mechanical loading.** The posterior surface of the tibia was loaded using 3-point bending analysis until failure to determine stiffness, stress, load at yield point, and ultimate load (Supplemental Fig. 3). Neither age nor dietary VA affected the yield point, ultimate load, or stress of bone when mechanically loaded (Supplemental Table 1). Stiffness was lower in middle-age VA-supplemented compared with VA-marginal rats ($P < 0.05$) but did not differ with VA in old rats.

**Whole bone mineral content.** Bones were weighed and ashed to determine the percentages of water, organic matrix, and ash, and the percentage of mineralized tissue was calculated (Fig. 2). Water percentage (Fig. 2A) was higher in young rats and slightly lower in the young VA-control group compared with VA-marginal and -supplemented groups ($P < 0.05$). Conversely, organic material was slightly higher in the young VA-control group (Fig. 2B). Neither ash (Fig. 2C) nor mineralized tissue (Fig. 2D) differed by age or dietary VA.

**Discussion**

There is essentially no information on VA dose-dependent changes on bone variables over the lifetime. In the present study,
we evaluated tibia from rats that had been fed 3 graded doses of VA, which were chosen to resemble the range of VA intakes of most humans. Our conditions thus ranged from VA-marginal to VA-supplemented, but we avoided both frank VA deficiency and toxicity (33,34,42,43). To our knowledge, this was a unique experiment with a high daily dose of VA. The results (Fig. 1) demonstrated that VA does travel to both the femur and tibia as animals grew and aged, bone mineral properties, bone parameters of bone quality, including bone dimensions over time could have contributed to the accumulation of VA. These results also supported the use of the tibia for bone analyses in our long-term study (Expt. 2).

In Expt. 2, by μCT analysis, some aspects of bone dimensions were altered by dietary VA in young rats (Table 1). Differences in VA intake (both marginal and supplemented) resulted in a reduction in cortical bone thickness and an increase in lumen size in young rats (Table 1). These results are consistent with current literature. It is noteworthy that young rats have been used in most previous studies on VA and bone (28). Although it may seem surprising that both VA-marginal and VA-supplemented young rats differed in a similar way from the VA-adequate group, there are other situations, e.g. fetal development, where similar outcomes have resulted from both VA deficiency and VA excess (46). Therefore, our VA-marginal and -supplemented diets may have resulted in too little and too much VA, respectively, for optimal bone formation during the phase of rapid growth. However, the long-term picture obtained by studying middle-age and old rats provides a new and unique look at the in vivo interaction of VA and bone. Despite the differences observed over a short period in young rats, there was no difference between the VA-supplemented compared with the control group for endosteal or periosteal radii, or cortical thickness, in middle-age and old rats (Table 1). Thus, with time the remodeling of the bone appears to have recovered in the VA-supplemented rats. On the other hand, the radial dimensions measured remained higher in VA-marginal compared with VA-supplemented rats. VA is known to be an important mediating factor in proper bone development (1,2).

### TABLE 1

Body weight and bone dimensions in tibia of young, middle-aged, and old rats fed diets differing in VA concentration for 2–3, 8–10, or 18–20 mo.\(^1,2\)

<table>
<thead>
<tr>
<th>Age</th>
<th>Diet</th>
<th>(n)</th>
<th>Body weight, g</th>
<th>Medullary area, (mm^2)</th>
<th>Endosteal radius, (mm)</th>
<th>Periosteal radius, (mm)</th>
<th>Cortical thickness, (mm)</th>
</tr>
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<tbody>
<tr>
<td>Young</td>
<td>Mar</td>
<td>6</td>
<td>378 ± 6.5(^{a})</td>
<td>2.58 ± 0.14(^{a})</td>
<td>0.89 ± 0.03(^{a})</td>
<td>1.50 ± 0.02(^{a})</td>
<td>0.49 ± 0.01(^{a})</td>
</tr>
<tr>
<td></td>
<td>Con</td>
<td>9</td>
<td>389 ± 5.4(^{a})</td>
<td>1.54 ± 0.06(^{a})</td>
<td>0.69 ± 0.02(^{a})</td>
<td>1.38 ± 0.01(^{a})</td>
<td>0.58 ± 0.02(^{a})</td>
</tr>
<tr>
<td></td>
<td>Sup</td>
<td>6</td>
<td>425 ± 43.2(^{a})</td>
<td>2.33 ± 0.11(^{a})</td>
<td>0.85 ± 0.02(^{a})</td>
<td>1.45 ± 0.02(^{a})</td>
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</tr>
<tr>
<td>Middle</td>
<td>Mar</td>
<td>7</td>
<td>584 ± 10.2(^{a})</td>
<td>3.14 ± 0.16(^{a})</td>
<td>0.94 ± 0.02(^{a})</td>
<td>1.74 ± 0.01(^{a})</td>
<td>0.60 ± 0.01(^{a})</td>
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<td>8</td>
<td>612 ± 18.0(^{a})</td>
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<td>641 ± 14.7(^{a})</td>
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<td>Con</td>
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<td>846 ± 30.5(^{ab})</td>
<td>4.34 ± 0.25(^{ab})</td>
<td>1.14 ± 0.04(^{b})</td>
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<td>901 ± 16.7(^{a})</td>
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<td>1.09 ± 0.03(^{a})</td>
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<td>0.55 ± 0.02(^{b})</td>
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Two-way ANOVA \(P\) values

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<th>Age</th>
<th>(P)</th>
<th>Interaction</th>
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\(^1\) Values are means ± SEM. Means in a column without a common letter differ, \(P < 0.05\).

\(^2\) Mar, marginal; Con, control; Sup, supplemented diet.

\(^3\) Illustrations of measurements can be found in Supplemental Figure 2.

### TABLE 2

Trabecular measurements in tibia of young, middle-aged, and old rats fed diets differing in VA concentration for 2–3, 8–10 or 18–20 mo.\(^1,2\)

<table>
<thead>
<tr>
<th>Age</th>
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<th>(n)</th>
<th>Trabecular, (\mu m/mm^2)</th>
<th>Trabecular thickness, (\mu m)</th>
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<td>Young</td>
<td>Mar</td>
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<td>3.7 ± 0.02(^a)</td>
<td>74 ± 2(^a)</td>
<td>657 ± 7(^d)</td>
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<td>3.4 ± 0.11(^b)</td>
<td>71 ± 3(^b)</td>
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<tr>
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<td>Sup</td>
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<td>69 ± 4(^d)</td>
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<tr>
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<td>Mar</td>
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<td>2.1 ± 0.05(^c)</td>
<td>147 ± 1(^b)</td>
<td>838 ± 4(^b)</td>
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<tr>
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<td>129 ± 2(^b)</td>
<td>807 ± 4(^c)</td>
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<td>Sup</td>
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<td>2.5 ± 0.12(^c)</td>
<td>130 ± 4(^b)</td>
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<tr>
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Two-way ANOVA \(P\) values

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\(^1\) Values are means ± SEM. Means in a column without a common letter differ, \(P < 0.05\).

\(^2\) Mar, marginal; Con, control; Sup, supplemented diet.

\(^3\) Illustrations of measurements can be found in Supplemental Figure 4.
marginal diet on cortical thickness, the overall bone shape as evaluated by endosteal and periosteal radii exhibited modest changes in middle-age rats that were lasting and significant by old age (Table 1). Both VA-marginal and VA-supplemented rats had mid-diaphyseal measurements that differed from those of control rats of the same age. These results indicate a difference in the shape of VA-supplemented rat bones by the time they reach middle age, which did not appear to be detrimental to the quality of the bone, because, as discussed below, all diet groups had similar responses to mechanical testing and similar mineralization.

The trabecular properties we measured changed with age and VA. The number and thickness of trabeculae as well as trabecular BMD showed significant differences in the VA-marginal group (Table 1). These results were consistent in both the middle-age and old groups and the variance in these measurements was quite small. These results suggest that further studies of the effect of VA and its metabolites, such as retinoic acid, on trabecular formation and maintenance would be of interest. An increase in the number of trabeculae in old VA-supplemented rats (Table 1), was not expected. However, this increase in trabecular bone might be explained by the differences in weight between the VA-marginal and VA-supplemented rats, or other unknown mechanisms, because the increase was normalized when trabecular number was adjusted for body weight. As seen in representative µCT reconstructions (Supplemental Fig. 4), long-term VA supplementation appears to have exerted a protective effect on trabecular bone loss associated with aging, while marginal VA status resulted in increased trabecular thickness and a slight increase in trabecular BMD. This suggests the potential for a biphasic response of bone to VA treatment, as has been suggested from the results of human epidemiological studies (6–9,11,16,18–30). Overall, our data suggest that dietary VA more dramatically affects trabecular bone than cortical bone in the tibia of rats. Although we were able to study only the tibia, other bone sites would also be of interest.

In our long-term study, multiple bone variables were evaluated, including mechanical and material properties testing of bone tissue and determination of bone composition. Diet did not affect the mechanical properties of the tibia tested by the 3-point bending test (Table 2) or bone composition as shown in Figure 2. These results imply that VA supplementation, at levels in excess of requirements but not high enough to produce hypervitaminosis A (33,34,42) did not have detrimental effects on bone loading capacity or strength. The larger variation within treatment groups for the materials testing analyses (Table 2) combined with the smaller sample size due to fewer bones being suitable for this analysis are limitations for our materials testing analyses. Although the expected outcome that VA supplementation would cause alterations in the mechanical properties of bone was not observed, the observance of no difference of VA-supplemented rats compared with control rats is also a potentially important point, because there is currently controversy regarding VA intakes above the RDA in human populations at risk for fracture.

Although we have illustrated our data without correction for body weight, we also analyzed our results after correcting for body weight, because weight can affect mechanical factors such as increased loading associated with changes to bone tissue, and increased weight can encourage mineralization and alter bone microarchitecture (47–49). The effects we reported for VA were similar both without and after normalization for body weight. Thus, the data presented reflect diet-dependent effects that are not due to age-related changes in body mass.

In summary, our study has revealed the potential for a VA-marginal diet to affect bone formation and architecture, especially of trabecular bone. Conversely, we did not observe differences in mechanical properties due to elevated dietary VA throughout the life time. The latter is regarded as an important finding, because VA has been regarded as a potential risk factor for reduced BMD and increased fracture risk in older humans. Our data show that higher intakes of VA, similar to those that have been proposed to reduce BMD and increase fracture risk in epidemiological studies (5–11,20), may not sufficiently alter bone properties related to strength to be detected in breaking strength assays. Our results more closely resemble the observational study reported by Sowers and Wallace (19), which investigated the relationship among current VA supplement use, serum retinol levels, radial bone mass, and fracture history in 246 postmenopausal women, 55–80 y of age. Although >36% of these women used a VA supplement, with 8% in excess of 2000 µg/d retinol, after controlling for relevant factors there was no significant relationship between VA supplement use or serum retinol and radial bone mass or fractures. In short-term studies,
Kawahara et al. (50) found no relationship between VA supplementation and bone turnover in men, and a recent large case-control study of patients treated with retinoid analogs that are in use for therapy did not observe increased risk of any hip, forearm, or spine fractures (51). Thus, the long-held notion that elevated dietary intake of VA is detrimental to bones requires further assessment. Animal models provide an ethical way to assess bone health with intakes of VA, which resemble those below and above levels currently recommended for human populations. Further investigations are needed to better understand VA signaling in bone tissue that affects bone modeling, microarchitecture, and mechanical properties across the lifetime, including growth, maintenance, and aging.

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Literature Cited

36. Reeves PG, Rossow KL, Lindlauf J. Development and testing of the AIN-93 purified diets for rodents: results on growth, kidney calcifica-